

Reduction of Potassium (K) Content in Strawberry Fruits through KNO₃ Management of Hydroponics

Md. Fuad Mondal^{1,2,3}, Md. Asaduzzaman⁴, Makoto Ueno¹, Mikiko Kawaguchi⁵, Shozo Yano⁶, Takuya Ban⁷, Hideyuki Tanaka¹ and Toshiki Asao^{1*}

¹Department of Agriculture, Faculty of Life and Environmental Science, Shimane University, Matsue 690-1102, Japan

⁴Olericulture Division, Horticulture Research Center, Bangladesh Agricultural Research Institute, Gazipur-1701, Bangladesh

⁵Faculty of Home Economics, Otsuma Women's University, Chiyoda-ku, Tokyo 102-8357, Japan

⁶Faculty of Medicine, Shimane University, Izumo 693-8501, Japan

⁷Faculty of Agriculture, Tokyo University of Agriculture and Technology, Fuchu 183-8509, Japan

The consumption of vegetables and fruits rich in potassium (K), such as melons and strawberries, is restricted in chronic kidney disease (CKD) patients. Therefore, we attempted to produce low-K strawberry fruits through management of a KNO₃ fertilizer in nutrient solution from anthesis to the harvest period. A general trend of decreasing K content in fruit was observed with the decrease of KNO₃ concentration in the nutrient solution. Among four strawberry cultivars, the fruit of the 'Toyonoka' exhibited a K reduction of about 64% when plants were grown in nutrient solution with KNO₃ at 1/16 of the normal level. Citric acid and ascorbic acid contents of 'Toyonoka' fruit were reduced with decreasing KNO₃ concentrations in the nutrient solution. Although the reduced NO_3^- of the nutrient solution was adjusted by using $Ca(NO_3)_2$ to obtain low-K strawberries, growth, yield, and quality did not vary with this adjustment. Compared with the typical level of K in strawberry fruit of 170 mg/100 g FW (Standard Tables of Food Composition in Japan, 2011), a 23.5% decrease (130 mg/100 g FW) in K was found in 1/32 level of KNO₃. The K contents of plant parts suggested that the low KNO₃ level was responsible for the low K absorption, which may have affected the translocation and accumulation of K into fruit. Therefore, 1/32 level of KNO₃ in nutrient solution lowers the fruit K content considerably.

Key Words: chronic kidney disease, *Fragaria* × *ananassa* Duch. 'Toyonoka', fruit potassium content, potassium nitrate, potassium translocation.

Introduction

Potassium (K) is a major electrolyte in the human body. It plays an important role in the contraction or relaxation of skeletal, smooth, and cardiac muscles, nerve impulse transmission, acid base equilibrium, enzymatic action, intracellular fluid tonicity, and renal function (Crawford and Harris, 2011; Russell, 2009). The average daily K intake is estimated to be 2000– 3900 mg·day⁻¹ (Choi and Ha, 2013; Kes, 2001; Putcha and Allon, 2007). About 77% of dietary K is excreted in the urine via the kidneys (Holbrook et al., 1984). A study found that higher K excretion is associated with a reduction in the risk of adverse renal outcomes (Smyth et al., 2014), but patients with chronic kidney disease (CKD) have limited ability to excrete K through the kidneys. As a result, the level of K increases in the blood serum. An elevated level of K in serum is called hyperkalemia, and is associated with an increased risk of chronic kidney disease progression and all-cause mortality in patients with chronic kidney disease (He et al., 2014). Mills et al. (2014) found that high dietary K was associated with an increased risk of cardiovascular disease among patients with CKD. In the United

²United Graduate School of Agricultural Sciences, Tottori University, Tottori 680-8553, Japan

³Faculty of Agriculture, Sylhet Agricultural University, Sylhet-3100, Bangladesh

Received; September 16, 2015. Accepted; March 6, 2016. First Published Online in J-STAGE on April 23, 2016.

^{*} Corresponding author (E-mail: asao@life.shimane-u.ac.jp).

States, hyperkalemia causes 5 deaths/1000 person-years in patients with CKD (US Renal Data System, 2002). The number of CKD patients is continuing to increase globally. In Japan, approximately 13.3 million people were predicted to have CKD in 2005, which was about 13% of the Japanese adult population (Imai et al., 2007). Therefore, considerable focus should be placed on preventing this disease. Potassium concentration in blood serum and CKD are positively correlated, so management of potassium intake by changing the diet is one approach to control CKD prevalence. He et al. (2014) suggested that the mortality rate of CKD patients can be reduced with a moderate reduction in dietary sodium and potassium intake. Within the daily diet, fruits such as melons and strawberries and leafy vegetables are rich in K, which is essential for maintaining good health. However, the consumption of these food stuffs by patients with CKD is restricted, which negatively impacts their quality of life (OOL). CKD patients are often restricted to 1500 mg K a day (Committee for Revision of Diet Therapy Guideline, 2007). Therefore, the development of fruits and vegetables with low K content would be beneficial for the above-mentioned patients to maintain their QOL.

Potassium is also an essential major nutrient for plant growth and development. Plants absorb more K than any other element, with the exception of N (Britto and Kronzucker, 2008; Szczerba et al., 2009). Voogt and Sonneveld (1997) found that with increases in tomato plant growth, K absorption increases to a relatively greater extent than that of other nutrients. Recently, low K content lettuce (Lactuca sativa L.) has been established by K fertilizer management (Ogawa et al., 2007). Hydroponic culture systems have become widely used because they allow greater control over the root zone environment than soil culture, which makes nutrient management easy based on the plant requirements. Hydroponic culture of strawberries has gained in popularity for the commercial production of strawberries (Koshikawa and Yasuda, 2003). Generally, greenhousecultured fresh strawberries have a high K content of 170 mg/100 g FW of fruit (Standard Tables of Food Composition in Japan, 2011). Reducing this level in strawberry fruit would provide a good option in the diet for CKD patients. Asao et al. (2013) reported that a reduction in KNO₃ in hydroponic solution could reduce the K concentration in melon fruit. Therefore, the objective of the present study was to investigate the impact of reduced K concentration in nutrient solution on the fruit K content of strawberries, while maintaining normal growth, yield, and other indicators of fruit quality.

Materials and Methods

1. Strawberry cultivars

Micro-propagated vigorous, disease-free, and similarly sized plantlets of four strawberry (*Fragaria* \times

ananassa Duch.) cultivars 'Toyonoka', 'Akihime', 'Saga-honoka', and 'Ai-berry' plantlets were used for this study.

2. Enshi nutrient solution

Strawberry plantlets were grown in 25% Enshi nutrient solution (pH 7.25 and EC 0.8 dS·m⁻¹). Fullstrength Enshi nutrient solution contains the following levels of salts per 1000 L of tap water: 950 g $Ca(NO_3)_2 \cdot 4H_2O; 810 \text{ g} \text{ KNO}_3; 500 \text{ g} \text{ MgSO}_4 \cdot 7H_2O;$ 155 g $NH_4H_2PO_4$; 3 g H_3BO_3 ; 2 g $ZnSO_4 \cdot 7H_2O$; 2 g MnSO₄·4H₂O; 0.05 g CuSO₄·5H₂O; 0.02 g Na₂MoO₄; and 25 g NaFe-EDTA (Hori, 1966). In the present study, we reduced the amount of KNO₃ while keeping the levels of other nutrients constant in order to produce strawberry fruit with low K content through hydroponic culture. Although the pH of the nutrient solutions was not varied, the EC decreased gradually with gradual decreasing KNO₃. The EC values of 25% nutrient solutions with 1/4, 1/8, 1/16, and 1/32 levels of KNO₃ were 0.70, 0.67, 0.65, and 0.48 dS \cdot m⁻¹, respectively. The EC and pH of tap water used for this experiment were $0.22 \text{ dS} \cdot \text{m}^{-1}$ and 8.18, respectively.

3. Expt. 1. Selection of suitable cultivars for the production of low-K strawberries from plants grown in nutrient solution with 1/1, 1/4, or 1/16 the normal level of KNO₃

In this experiment, the above-mentioned four strawberry cultivars were evaluated for low K content in strawberry fruit using a hydroponic system inside a greenhouse $(20 \text{ m} \times 5 \text{ m})$ of the Experimental Research Center for Biological Resources Science, Shimane University. The experiment was conducted from September 2009 to June 2010. The greenhouse had no heating system and its top windows opened at temperatures over 25°C. Micro-propagated strawberry plantlets were transferred into cell trays ($48 \text{ cm} \times 24 \text{ cm} \times 4 \text{ cm}$, 72 cells/tray) with vermiculite substrate on 14 September, 2009, and kept there until 6 October, 2009, under controlled growth chamber conditions at 20°C/ 15°C (day/night), 60% relative humidity (RH), fluorescent light with intensity of 145 μ mol \cdot m⁻²·s⁻¹, and a 12 h photoperiod for the formation of new roots and leaves. Twenty-five-percent standard Enshi nutrient solution was used for fertilization. Then, the cell trays were transferred into the greenhouse and kept there until 26 October, 2009. On 27 October, 2009, 6 four to five leafstage strawberry plantlets were transplanted into plastic containers ($20 \text{ cm} \times 54 \text{ cm} \times 34 \text{ cm}$) with 50 L of 25% nutrient solution. Before transplanting, vermiculite adhering to the strawberry plantlet roots were washed gently by tap water in a bucket. A urethane foam block $(23 \text{ mm} \times 23 \text{ mm} \times 27 \text{ mm})$ was used to hold the plants tightly with a floating board on the nutrient solution. Nutrient solutions were circulated for 24 h using pumps (KP-101; Koshin, Kyoto, Japan) The culture solutions

were renewed every two weeks throughout the entire growth period. Reduction of KNO₃ in the culture solution was started after the first anthesis of all plants. 25% standard nutrient solution with 1/1. 1/4. or 1/16 the normal level of KNO₃ was used during plant growth. In this experiment, the total number of strawberry plants (4 cultivars \times 3 level of KNO₃ \times 3 replications \times 6 plantlets per treatment) was 216. Pollination was facilitated using a soft brush at two-day intervals. Fruit were harvested when they became about 80% red in color. At each harvest, the fresh weight of fruit was recorded and used for the final yield calculation. At final harvest on 14 June, 2010, the number of leaves and root length were recorded. Then, strawberry plant parts were separated into leaf crowns and roots, and dried in a constant temperature oven (DKN 812; Yamato Scientific Co., Ltd., Japan) for 72 h at 80°C. When the dry matter reached a constant weight, the dry weights of different plant parts were measured.

 Expt. 2. Culture of F. × ananassa Duch. 'Toyonoka' in nutrient solutions with 1/1, 1/8, and 1/16 the normal level of KNO₃

This experiment was carried out in a controlled room of the plant factory support research lab of the Experimental Research Center for Biological Resources Science, Shimane University. From the earlier experiment, 'Toyonoka' was selected for this experiment. Plantlets were kept for 7 weeks from 29 October, 2010 to 19 December, 2010 in controlled conditions as maintained in Expt. 1 for flower bud induction. Then, five bud-formed plantlets were planted in a hydroponic culture bed using four urethane blocks as support filled with 50 L of nutrient solution at $25^{\circ}C/20^{\circ}C$ (day/night). 60% RH, under fluorescent light with intensity of $145 \,\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ and a $12 \,\text{h}$ photoperiod on 20 December, 2010. Three beds were used for one treatment connected with a 150-L nutrient solution tank (Fig. 1). Nutrient solutions were recycled at 55/5 min (recycle/stop) using an automatic pump. Treatment included 25% standard Enshi nutrient solution with 1/1, 1/8, and 1/16 the normal level of KNO₃. In this experiment, the total number of strawberry plants (3 level of $KNO_3 \times 3$ replications $\times 5$ plantlets per treatment) was 45. Culture solution was renewed biweekly. Pollination was performed using the brush at two-day intervals. At maturity, the fresh weight of fruit was recorded and used for a final yield comparison among the treatments. At final harvest on 14 June, 2011, plant growth parameters were measured and dry weights of the different plant parts were measured after oven drying of the samples, as described for Expt. 1.



Fig. 1. Hydroponic system under a controlled environment where strawberry plants were cultured in bed with 50 L nutrient solution connected to a 150-L nutrient solution tank.

5. Expt. 3. Production of low-K strawberry fruits using culture solutions with 1/8, 1/16, and 1/32 the normal level of KNO₃ with or without adjustment of NO₃⁻ supply by Ca(NO₃)₂

This experiment was also carried out in a controlled room of the plant factory. On 4 April, 2012, 'Toyonoka' plantlets were kept for bud formation under controlled environmental conditions as described for Expt. 2. Five bud-formed plantlets were planted in a hydroponic culture bed on 15 June, 2012. Three beds were used for one treatment connected with a 150-L nutrient solution tank. Five types of nutrient solution were used as 25% standard nutrient solution with (i) 1/8 KNO₃, (ii) 1/16 KNO_3 , (iii) 1/32 KNO_3 , (iv) 1/16 KNO_3 with $Ca(NO_3)_2$ adjusted to the same NO₃⁻ levels as 1/8 KNO₃, and (v) 1/32 KNO₃ with Ca(NO₃)₂ adjusted to the same NO₃⁻ levels as 1/8 KNO₃. EC and the pH values of these nutrient solutions were 0.67, 0.65, 0.48, 0.54, and 0.53 dS·m⁻¹ and 7.57, 7.60, 7.33, 7.55, and 7.53, respectively. In number (iv) and (v) types of nutrient solution, Ca(NO₃)₂ was used to recover the effects of reduced NO₃⁻ of KNO₃. Culture solutions were renewed biweekly. In this experiment, the total number of plantlets (5 level of $KNO_3 \times 3$ replications \times 5 plantlets per treatment) was 75. Pollination and fruit harvest were performed as mentioned for the earlier experiments. Harvested fruit were divided into three categories on the basis of pollination date, namely, stage 1 or early stage (fruit collected from 18/6/2012 to 22/6/2012

as the date of pollination), stage 2 or mid-stage (fruit collected from 16/7/2012 to 20/7/2012 as the date of pollination), and stage 3 or late stage (fruit collected from 13/8/2012 to 13/8/2012 as the date of pollination). On 28 September, 2012, growth parameters were measured at the final harvest. The dry weights of different plant parts were measured after oven drying of the samples as described for Expt. 1.

6. Measurement of soluble solid, titratable acid, and ascorbic acid contents of strawberry fruit

After harvest, the fruit was composited and frozen at -30°C for subsequent analysis of soluble solid, titratable acid, and ascorbic acid contents. Soluble solid content was measured by placing a fruit juice sample into a pocket digital refractometer (PAL-1; Atago Ltd., Tokyo, Japan). Titratable acid content was determined by diluting a 2 mL aliquot of strawberry juice to 10 mL with 8 mL of distilled water and adding 2-3 drops of phenolphthalein, followed by adjustment of the pH to 8.2 using 0.1 N (w/v) NaOH. The quantity of NaOH (mL) and the amount for appropriate acidity were converted into citric acidity (%). The ascorbic acid content was measured using 2,4-dinitrophenylhydrazine (DNP) colorimetry. Strawberry fruit juice (0.5 mL) was placed in a 50-mL test tube, to which 0.5 mL of 10% metaphosphoric acid solution, 1 mL of distilled water, 1 mL of 0.03% 2,6-dichlorophenol-indophenol (DCP), 2 mL of thiourea, and 1 mL of 2,4-DNP were added, followed by 3 h of incubation at 37°C in a water bath (BW400; Yamato Scientific Co., Ltd., Tokyo, Japan). After incubation, 5 mL of 85% H₂SO₄ was added while the samples were kept cool with iced water. After 30 min of cooling, the ascorbic acid content was measured at 520 nm using a spectrophotometer (U-2900; Hitachi High Technologies Corporation, Tokyo, Japan).

7. Determination of K and other nutrient concentrations in strawberry fruit

In order to measure the concentrations of K including Ca, Mg, Fe, and Na, fruit were kept in a 15-mL plastic test tube and left on ice for subsequent analysis of minerals. On the day of analysis, strawberry samples were taken out of the freezer for thawing. Then, the fresh weight of each fruit sample (8-10 g) was measured (Excellence XS Analytical Balance, XS204DRV; Mettler Toledo, Greifensee, Switzerland). Each sample was then placed in a 250-mL plastic bottle containing 200 mL of 1% HCl. The samples were next shaken in a Bio-Shaker (Bio-Shaker BR-43FL; TAITEC, Koshigaya, Japan) for 30 min at 150 rpm in order to liquefy the fruit flesh completely. The dissolved fruit samples were then filtrated (Advantec Grade No. 131, 185 mm; Advantec, Tokyo, Japan) and analyzed for the above minerals with a polarized Zeeman Atomic Absorption Spectrophotometer (Z-2310; Hitachi High Technologies Corporation).

8. Determination of K and other nutrient concentrations in strawberry plant parts

When the dry matter reached a constant weight, it was ground into powder with a mixer machine (MX-X53; National, Tokyo, Japan). Samples weighing 0.5 g were mixed with 8 mL of 97% HNO₃ and digested using a microwave sample preparation system (ETHOS1; Milestone S.r.l., Bergamo, Italy). After digestion, samples were brought up to a total of 50 mL in a volumetric flask by the addition of distilled water and then filtered with qualitative filter paper (Grade No. 131). The filtered sample solutions were analyzed for K, Ca, Mg, Fe, and Na using a Zeeman Atomic Absorption Spectrophotometer.

9. Statistical analysis

Analysis of variance was performed to test for significant effects of different KNO₃ levels in the nutrient solution on plant growth, fruit yield, and the quality and nutrients in parts of strawberry plants in all three experiments. Means were compared by Duncan's multiple range test (Mstat-C statistical software; Mstat Director, Department of crop and soil sciences, Michigan state University, USA) and Tukey's multiple range test (Statcel 2 Software; OMS Publications, Tokorozawa, Japan) at P < 0.05 and P < 0.01.

Results

1. Effects of reduced KNO₃ on the growth, yield, and fruit quality of four strawberry cultivars

It was cleared that strawberry cultivars significantly influenced the plant growth parameters while KNO₃ levels did not show any significant effects on those parameters (Table 1). Higher numbers of leaves were recorded for the 'Toyonoka' cultivar when plants were grown in the nutrient solution with 1/1 and 1/4 levels of KNO₃ and in 'Saga-honoka' with 1/1 and 1/16 levels of KNO3. In terms of root length, longest and smallest roots were found in 'Toyonoka' with a 1/4 level of KNO₃ and 'Ai-berry' with 1/1 level of KNO₃. Among the four strawberry cultivars, higher and lower amounts of leaf dry matter were founded in 'Saga-honoka' and 'Ai-berry', respectively. From the interaction it was founded that 'Toyonoka' with a 1/16 level of KNO₃, 'Akihime' with a 1/1 level of KNO₃, and 'Ai-berry' with all levels of KNO3 showed reductions in their leaf dry matter.

The results showed that the numbers of strawberry fruits per plants were influenced by the cultivars, but not by KNO_3 levels (Table 2). Among the four cultivars, 'Toyonoka' with 1/1 level of KNO_3 produced the greatest number of fruits per plant, whereas the lowest number was found in 'Ai-berry' with a 1/16 level of KNO_3 . Neither the cultivars nor the KNO_3 levels influenced the fruit weight per plant. However, 'Toyonoka' with a 1/1 and 'Akihime' with a 1/4 level of KNO_3 produced the greatest amount of fruit per plant. Larger fruit

Cr. 1 10	KNO ₃ ^z	T C I	v	Root length			Dry w	veight (g)	
Strawberry cultivars	treatments	Leaf number	'S'	(cm)		leave	es	crov	vn
Toyonoka		40.7	a ^x	47.7	a	26.0	ab	1.8	a
Akihime		23.4	b	38.5	а	25.9	ab	3.1	а
Saga-honoka		41.2	а	46.9	а	40.3	а	2.2	а
Ai-berry		25.5	ab	34.3	а	21.7	b	3.1	а
Significant level			**		NS		**		NS
	1/1	33.4	a	39.1	a	28.9	a	2.6	a
	1/4	34.5	а	43.4	а	30.0	а	2.6	а
	1/16	30.3	а	42.9	а	26.3	a	2.5	а
Significant level			NS		NS		NS		NS
Toyonoka	1/1	44.6	а	43.6	abcd	29.3	bc	1.6	f
	1/4	44.9	а	51.6	а	29.4	bc	1.8	ef
	1/16	32.7	abc	48.1	ab	19.4	c	1.9	def
Akihime	1/1	20.7	c	34.3	de	22.5	c	2.9	abcde
	1/4	25.8	c	40.8	bcde	28.5	bc	3.1	abcd
	1/16	23.7	c	40.3	bcde	26.9	bc	3.3	ab
Saga-honoka	1/1	41.9	а	45.4	abc	42.6	a	2.6	abcdef
	1/4	39.3	ab	45.4	abc	38.2	ab	2.1	bcdef
	1/16	42.4	а	49.8	ab	40.0	ab	2.0	cdef
Ai-berry	1/1	26.4	с	33.2	e	21.2	c	3.4	а
	1/4	27.9	bc	36.2	cde	23.9	c	3.2	abc
	1/16	22.3	c	33.6	de	19.8	c	2.7	abcdef
Significant level			*		*		*		*

Table 1. Effects of reduced KNO₃ on the growth of four strawberry cultivars grown in hydroponic nutrient solution.

^z KNO₃ concentration in 25% standard Enshi nutrient solution.

^y Parameters were measured on a per plant basis.

^x Means within a column followed by different letters are significantly different according to Duncan's multiple range test at P < 0.05 (n=18).

NS: not significant, * Significant at P < 0.05, ** Significant at P < 0.01.

were found for ta he 'Ai-berry' cultivar at all levels of KNO₃.

Fruit quality was also affected by the strawberry cultivars and reduced levels of KNO₃. Among the cultivars and KNO₃ level 'Toyonoka' with a 1/1 level of KNO₃ produced strawberries with higher ascorbic acid contents while it was lower in other cultivars with 1/16 of KNO₃ (Table 2). Although the cultivars and KNO₃ level did not significantly affect the soluble solid contents, 'Saga-honoka' fruit with 1/1 level of KNO₃ and 'Aiberry' fruit with 1/16 KNO₃ produced the highest and lowest soluble solid contents, respectively. With the reduction in KNO₃ in the culture solution, K content in all cultivars decreased gradually (Table 2). Among the four strawberry cultivars, the lowest K was measured in 'Toyonoka' with 1/16 KNO₃. Compared with the control (1/1 level of KNO₃), the reduction was approximately 64%.

2. Culture of 'Toyonoka' in nutrient solution with 1/1, 1/8, and 1/16 levels of KNO₃

When strawberry plants were grown in nutrient solution with different reductions in the levels of KNO_3 in a controlled environment, the plant growth parameters significantly varied among the treatments (Table 3). Compared with the 1/1 level of KNO₃, leaf numbers were significantly reduced with KNO₃ at 1/16 the normal level. Root length was also affected by the reduced level of KNO₃. Compared with 1/1 level of KNO₃, smaller roots were found in plants grown at KNO₃ 1/8 and 1/16 levels. Crown diameters of strawberry plants also decreased with the reduction in KNO₃ level in the culture solution. It was shown that, compared with the control, both 1/8 and 1/16 levels of KNO₃ reduced the fresh weight of leaves and crowns and the dry weight of leaves, crown, and roots.

Reduced KNO₃ concentration had significant effects on the yield and fruit quality of strawberries (Table 4; Fig. 2). Compared with the control, total fruit weight per plant was higher with KNO₃ at 1/8. Although the fruit numbers did not differ among the treatments, larger fruit were found at a 1/16 level of KNO₃. Fruit quality characteristics, such as citric acidity (%) and ascorbic acid content, were reduced with reductions in KNO₃ concentration. Compared with those at the standard KNO₃ concentration in the nutrient solution, soluble solids did not vary among the treatments.

Fruit K concentration was greatly influenced by the

Strawberry cultivars	KNO ₃ ^z	No. of fruits/plan	ıt	Fruit weight (g/plant)	Single frui weight (g	it)	Soluble solids (%)	;	Ascorbi acid (ppm)	c	Fruit K (mg/100 g)
Toyonoka		27.0	a ^y	201.0	а	7.6	a	8.2	a	825.0	а	224.0	ab
Akihime		25.1	а	200.8	а	7.5	а	8.8	a	466.6	b	274.8	b
Saga-honoka		19.8	ab	148.0	а	7.4	а	8.7	а	494.3	b	235.5	ab
Ai-berry		14.7	b	151.9	а	10.8	а	7.4	а	448.7	b	198.8	b
Significant level			**		NS		NS		NS		**		**
	1/1	23.2	а	175.9	а	7.8	а	9.1	a	631.1	а	304.1	а
	1/4	21.9	а	183.4	а	8.6	а	8.1	а	583.5	ab	232.3	b
	1/16	20.4	а	166.9	а	8.6	а	7.6	а	444.6	b	163.4	c
Significant level			NS		NS		NS		NS		**		**
Toyonoka	1/1	29.8	а	217.7	а	7.3	b	9.1	ab	1038.1	а	311.1	ab
	1/4	27.8	ab	205.6	ab	7.5	b	8.2	abc	963.3	а	248.3	cde
	1/16	23.6	abcd	179.7	abcd	7.8	b	7.5	bc	406.1	b	112.7	h
Akihime	1/1	26.4	abc	186.5	abcd	6.8	b	9.3	ab	480.9	b	353.6	а
	1/4	22.7	abcde	214.5	а	7.6	b	9.0	ab	462.0	b	267.8	bcd
	1/16	26.1	abc	201.6	abc	8.1	b	8.2	abc	457.0	b	202.9	ef
Saga-honoka	1/1	20.7	bcdef	145.1	d	7.3	b	10.1	а	526.3	b	285.4	bc
	1/4	19.8	cdefg	154.5	cd	7.7	b	7.8	bc	471.6	b	229.9	def
	1/16	18.8	defg	144.1	d	7.3	b	8.5	abc	484.9	b	191.2	fg
Ai-berry	1/1	16.0	efg	154.4	cd	9.6	ab	8.3	abc	479.0	b	266.4	bcd
	1/4	14.8	fg	159.1	bcd	11.3	а	7.7	bc	436.9	b	183.0	fg
	1/16	13.2	g	142.2	d	11.3	а	6.3	с	430.2	b	146.7	gh
Significant level			*		*		*		*		**		*

 Table 2. Effects of reduced KNO3 on the yield and quality of strawberry fruit from four strawberry cultivars grown in hydroponic nutrient solution.

^z KNO₃ concentration in 25% standard Enshi nutrient solution.

^y Means within a column followed by different letters are significantly different according to Duncan's multiple range test at P < 0.05 (n=18). NS: not significant, * Significant at P < 0.05, ** Significant at P < 0.01.

 Table 3. Effects of reduced KNO₃ on the growth of strawberries 'Toyonoka' grown in renewed nutrient solution under controlled environmental conditions.

	N. C			Leaf		Crown	Fresh	weight (g)	Dry weight (g)			
KNO ₃ ^z	No. of leaves ^y		length (cm)	width (cm)	length (cm)	diameter (cm)	leaves	crown	leaves	crown	roots	
1/1	60.8	a ^x	11.9 ab	16.2 a	45.0 a	1.3 a	255.7 a	21.5 a	44.5 a	1.5 a	1.4 a	
1/8	51.9	ab	9.6 b	16.6 a	40.8 b	1.3 a	136.1 b	10.7 b	32.6 b	0.3 b	0.6 ab	
1/16	44.7	b	12.5 a	16.4 a	37.5 b	1.0 b	91.5 c	9.9 c	26.7 b	0.2 b	0.4 b	
		*	*	NS	*	*	*	*	*	*	*	

^z KNO₃ concentration in 25% standard Enshi nutrient solution.

^y Parameters were measured on a per plant basis.

^x Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P < 0.05 (n=15).

NS: not significant, * Significant at P < 0.05.

reduction in KNO₃ concentration in the culture solution (Fig. 3). Compared with the 1/1 level of KNO₃, K content was decreased by 43% to 54% at 1/8 and 1/16 KNO₃, respectively. When the K concentrations of plant dry matter from different treatments were analyzed, significant differences were observed (Figs. 4–6). With the reduction in KNO₃ concentration in the culture solution, the K contents in leaves, crowns, and roots decreased gradually. Compared with that in the control, K

content was reduced by 85% and 88% in leaves, 76% and 84% in crowns, and 86% and 89% in roots at 1/8 and 1/16 level of KNO₃, respectively.

3. Production of low-K strawberries through adjustment using Ca(NO₃)₂ in culture solution from 1/16 and 1/32 levels of KNO₃ to a 1/8 level of KNO₃

Compared to a 1/32 level KNO₃, leaves, crowns, and root dry matter of strawberry plants did not increased at

KNO3 ^z	KNO ₃ ^z Fruit weight (g/plant)		Fruit no./plan	t	Single fruit weight (g)		Soluble solid (%)	Soluble solids (%)		Citric acidity (%)		d
1/1	331.3	b^{y}	50.9	а	6.5	b	7.8	а	0.39	а	460.5	а
1/8	431.0	а	57.3	а	7.5	а	8.1	а	0.28	b	435.1	b
1/16	379.9	b	48.6	а	7.9	а	7.7	а	0.23	b	427.9	b
		*		NS		*		NS		*		*

 Table 4. Effects of reduced KNO3 on the yield and fruit quality of strawberries ('Toyonoka') grown in nutrient solution under controlled environmental conditions.

^z KNO₃ concentration in 25% standard Enshi nutrient solution.

^y Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P < 0.05 (n=15).

NS: not significant, * Significant at P < 0.05.



Fig. 2. Strawberry fruits in a hydroponic system under a controlled environment.



KNO3 concentration in 25% standard nutrient solution

Fig. 3. Effects of reduced KNO₃ levels in culture solution on strawberry 'Toyonoka' fruit potassium content. Data are presented as means \pm SE (n = 15). Different letters indicate significant differences between the treatments (P < 0.05).

a 1/32 level of KNO₃ with adjustment of the NO₃⁻ supply by Ca(NO₃)₂ (Table 5). There were no significant effects of Ca(NO₃)₂ adjustment on the yield and fruit quality, in terms of soluble solids, citric acidity, NO₃⁻, and ascorbic acid concentration (Table 6). Compared with a 1/8 level of KNO₃, all levels of reduced KNO₃ in the nutrient solution reduced fruit K concentration in all three stages of harvested fruit (Table 7). K concentration in the strawberry fruits did not differ significantly in Ca(NO₃)₂ adjusted 1/32 level KNO₃ compared with a



Fig. 4. Effects of reduced KNO₃ levels in culture solution on the potassium absorption by strawberry 'Toyonoka' leaves. Data are presented as means \pm SE (n = 15). Different letters indicate significant differences between the treatments (P < 0.05).



Fig. 5. Effects of reduced KNO₃ levels in culture solution on the potassium absorption by strawberry 'Toyonoka' crown. Data are presented as means \pm SE (n = 15). Different letters indicate significant differences between the treatments (P < 0.05).





Table 5. Effects of $Ca(NO_3)_2$ adjustment for reduced NO_3^- of KNO_3 on the growth of strawberry plants.

KNO Z No. of		Leaf length		Leaf width	Leaf width		Root length		Dry weight (g)					
KNO ₃ -	leaves ^y		(cm)		(cm)		(cm)		leaves		crown		roots	
1/8	19.1	a ^w	241.8	a	166.9	а	501.6	b	12.8	a	1.5	a	4.3	а
1/16	14.3	b	222.3	ab	168.1	а	557.0	ab	12.5	а	1.2	ab	4.5	a
1/32	14.9	b	199.9	ab	145.7	b	556.0	ab	8.7	b	0.9	b	3.4	b
1/16 (adjusted)x	18.1	ab	219.3	ab	157.0	ab	600.3	ab	11.0	а	1.3	ab	4.0	ab
1/32 (adjusted)x	14.7	b	191.0	b	158.0	ab	546.5	ab	9.3	ab	1.0	ab	3.8	ab
		*		*		*		*		*		*		*

^z 25% standard Enshi solution with KNO₃ conc. of 1/8, 1/16, and 1/32.

^y Parameters were measured on a per plant basis.

^x 1/16 and 1/32 KNO₃ with Ca(NO₃)₂ adjustment to 1/8 KNO₃.

^w Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P<0.05 (n=15).

* Significant at P<0.05.

Table 6.	Effects of Ca(NO ₃) ₂ adjustment on reduced NO	$_{2}^{-}$ of KNO ₂ on the yield and fruit quality of strawberry fr	uits.
14010 01	Enterts of Cu(103)2 unjubilitent on reduced 100	j of fill (of on the field and fight quantif of bud (of only in	careo.

KNO3 ^z	Fruit weight (g/plant)	No. of fruits/plant	Soluble solids (%)	Citric acidity (%)	NO ₃ ⁻ (ppm)	Ascorbic acid (ppm)
1/8	160.8 a ^y	28.3 a	8.4 a	0.32 a	589.0 a	237.1 a
1/16	172.0 a	25.0 a	8.4 a	0.32 a	759.3 a	233.1 а
1/32	169.0 a	26.1 a	7.7 a	0.29 a	546.9 a	250.1 a
1/16 (adjusted)x	165.7 a	27.1 a	8.8 a	0.32 a	851.0 a	244.2 a
1/32 (adjusted)x	167.9 a	25.4 a	7.5 a	0.29 a	754.6 a	255.4 a
	NS	NS	NS	NS	NS	NS

 z 25% standard Enshi solution with KNO3 conc. of 1/8, 1/16, and 1/32.

^y Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P<0.05 (n=15).

 x 1/16 and 1/32 KNO₃ with Ca(NO₃)₂ adjusted to 1/8 KNO₃.

NS: not significant.

	K (mg/100 g)			C	Ca (mg/100	0 g)	Mg	(mg/100	g)	Na	u (mg/100) g)	Fe	(mg/100	g)
KNO_3^2	S1 ^y	S2	S3	S1	S2	S3	S1	S2	S3	S1	S2	S3	S1	S2	S3
1/8	237 a ^x	199 a	183 a	85 a	1 89 ab	84 b	29 a	31 a	23 a	6.4 ab	6.8 a	3.4 a	2.4 a	2.5 a	1.9 a
1/16	181 b	160 b	161 b	77 a	1 73 b	83 b	31 a	38 a	25 a	8.0 a	5.1 b	3.1 a	1.9 a	1.9 a	1.7 a
1/32	168 bc	146 bc	130 c	70 a	1 78 b	90 ab	31 a	33 a	25 a	4.9 b	5.5 b	3.4 a	1.7 a	1.7 a	1.3 a
1/16 (adjusted)w	148 c	131 c	156 b	69 a	u 97 a	101 a	27 a	29 a	27 a	5.6 ab	5.6 b	3.7 a	1.6 a	1.3 a	1.6 a
1/32 (adjusted)w	149 c	135 c	126 c	79 a	104 a	97 a	28 a	32 a	27 a	5.2 b	7.3 a	3.5 a	1.6 a	1.4 a	1.4 a
	*	*	*	Ν	S *	*	NS	NS	NS	*	*	NS	NS	NS	NS

Table 7. Effects of Ca(NO₃)₂ adjustment on reduced NO₃⁻ of KNO₃ the mineral contents of strawberry fruits.

^z 25% standard Enshi solution with KNO₃ conc. of 1/8, 1/16, and1/32.

^y S1=stage 1 (fruit collected from 18/6/2012 to 22/6/2012 as the date of pollination), S2=stage 2 (fruit collected from 16/7/2012 to 20/7/2012 as the date of pollination), S3=stage 3 (fruit collected from 13/8/2012 to 13/8/2012 as the date of pollination).

^x Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P < 0.05 (n=15).

w 1/16 and 1/32 KNO₃ with Ca(NO₃)₂ adjusted to 1/8 KNO₃.

NS: not significant, * Significant at P < 0.05.

1/32 level of KNO₃. In terms of the calcium content of strawberry fruit, stage 1 fruit did not show any significant variation; however, in stage 2 and stage 3, calcium content increased significantly with Ca(NO₃)₂ adjustment for both 1/16 and 1/32 levels of KNO₃. Na content was increased in stage 2 fruit with Ca(NO₃)₂ adjustment of 1/32 level of KNO₃. Nutrient contents of strawberry leaves significantly varied among treatments of reduced KNO₃ and Ca(NO₃)₂-adjusted KNO₃ (Table 8). Com-

pared with a 1/8 level of KNO₃, leaf K contents were decreased by about 37%, 56%, 46%, and 60% in plants grown with treatments of 1/16 and 1/32 levels of KNO₃, and Ca(NO₃)₂ adjustment of 1/16 and 1/32 KNO₃, respectively. Other nutrients in leaves, such as Ca, Na, and Fe, decreased with decreases in KNO₃ concentration in the nutrient solution. However, when the reduced NO₃⁻ was adjusted by using Ca(NO₃)₂, Ca content of leaves was increased with 1/16 level of

KNO ₃ ^z	K (ppm) (×100)	Ca (ppm) (×10)	Mg (ppm) (×10)	Na (ppm) (×10)	Fe (ppm) (×1)
1/8	3.15 a ^x	5.19 a	0.43 b	0.43 a	1.36 a
1/16	1.98 b	4.69 b	0.53 a	0.40 ab	1.28 b
1/32	1.36 c	4.64 b	0.51 a	0.36 b	1.10 c
1/16 (adjusted) ^y	1.68 b	5.18 a	0.44 b	0.38 b	1.19 c
1/32 (adjusted) ^y	1.23 c	4.48 b	0.48 ab	0.33 b	1.16 c
	*	*	*	*	*

Table 8. Effects of $Ca(NO_3)_2$ adjustment on reduced NO_3^- of KNO_3 on the mineral contents of strawberry leaves.

^z 25% standard Enshi solution with KNO₃ conc. of 1/8, 1/16, and 1/32.

^y 1/16 and 1/32KNO₃ with Ca(NO₃)₂ adjusted to 1/8 KNO₃.

^x Means within a column followed by different letters are significantly different according to Tukey's multiple range test at P < 0.05 (n=15).

* Significant at P<0.05.

 KNO_3 . The Ca(NO_3)₂ adjustment of reduced KNO_3 did not increase the Na or Fe contents of leaves. Mg content increased with reduced KNO_3 in the nutrient solution.

Discussion

Among four strawberry cultivars, 'Toyonoka' and 'Ai-berry' plants exhibited reductions in the dry matter contents with 1/16 level of KNO₃ in the nutrient solution (Table 1). In another experiment, it was found that the dry matter contents of leaves, crowns, and roots significantly decreased with decreases in KNO₃ fertilizer in the nutrient solution (Table 3). This growth reduction was attributed to reductions in leaf number. Potassium is the cationic nutrient required at the highest level by plants, constituting up to 10% of plant dry matter (Leigh and Wyn Jones, 1984). This reduction in biomass because of K⁺ deficiency is often accomplished by a reduction in leaf area (Jordan-Meille and Pellerin, 2004). Moreover, nitrate is generally the preferred source for crop growth (Miller and Cramer, 2004). Besides the KNO₃ reduction in the nutrient solution, the difference in cultivars also has an effect on the K uptake and utilization for the normal growth of plants. Woodend and Glass (1993) studied the uptake and utilization of K in wheat under conditions of K stress and found some variation among the varieties and genotypes.

Compared with the control (full strength of KNO₃), 'Ai-berry' exhibited decreases in fruit yield by about 8% with 1/16 KNO₃ in the nutrient solution (Table 2). It has been reported that K deficiency is related to reduced stomatal conductance, thereby impairing CO₂ fixation, which disrupts the conversion of light energy to chemical energy, and the phloem export of photosynthates from source to sink (Cakmak, 2005). In the case of maize and wheat production, when an insufficient level of K in soil was recovered using K fertilizer, the yield of those crops was increased (Ebelhar and Varsa, 2000; Singh and Sharma, 2001). However, when 'Toyonoka' was grown in a plant factory with a 1/8level of KNO₃, its yield improved compared with that of the control (Table 4). This improved yield was attributable to a higher number of fruit. Luxurious vegetative growth was found in plants grown with control nutrient solution, leading to the lowest yield. Egilla et al. (2001) reported that an adequate amount of K can enhance the total dry matter accumulation of crop plants in comparison to lower K concentration.

The soluble solid contents of 'Saga-honoka' fruits were reduced with reductions of KNO₃ in the culture solution. In terms of ascorbic acid content, a similar trend was found in 'Toyonoka' cultivars (Table 2). Researchers have generally suggested that adequate K nutrition increases yield, fruit size, soluble solids, and ascorbic acid concentration, as well as improving fruit color, and increasing the shelf life and shipping quality of many horticultural crops (Kanai et al., 2007; Lester et al., 2005, 2006). Reduced KNO3 in the nutrient solution with Ca(NO₃)₂ adjustment did not show any effects on the citric acid and ascorbic acid contents of strawberry fruit. Some studies have shown that there was little or no change in fruit quality with K fertilization in apples, cucumbers, mangoes, bell peppers, strawberries, and watermelons (Albregts et al., 1996; Hassanloui et al., 2004; Hochmuth et al., 1994; Locascio and 2002; Perkins-Veazie et al., Hochmuth, 2003; Rebolledo-Martinez et al., 2008; Umamaheswarappa and Krishnappa, 2004).

It was found that simple management with K fertilizer in the nutrient solution could reduce the level of K in strawberry fruit in all of the examined strawberry cultivars. Among these cultivars, 'Toyonoka' plants with a 1/16 level of KNO₃ exhibited a reduction of K of 64% in fruit compared with the control (Table 2). In two other independent experiments, the same level of KNO₃ reduced K by 54% and 32% in strawberry fruit (Fig. 3; Table 6). A general trend of decreasing K content in fruit was observed with decreasing KNO₃ concentrations in the nutrient solution. These results are in accordance with the study of Asao et al. (2013) in which the K of melon fruit decreased with decreasing KNO₃ levels in the nutrient solution. The reduction of K in the fruit may have been due to its low absorption by the

plants from the nutrient solution, low partitioning in different plant parts, and finally low accumulation in fruit. The total amount of K absorbed by the crops during the growing season depends upon the crop species being grown, the amount of native K⁺ in the soil, the amount of K⁺ applied to the fertilizer, its availability in the soil, the environmental conditions during the growing season, and management practices (Mullins and Burmester, 1998). In other research, it was found that K uptake also depends on plant-related factors, including genetics and the developmental stage, such as vegetative versus reproductive stages (Rengel and Damon, 2008). Pettersson and Jensen (1983) found differences among barley genotypes in terms of K influx and efflux, transport to shoots, and K use efficiency (dry matter produced per amount of absorbed K). On the other hand, the reduced level of KNO₃ also reduced the NO₃⁻ in the nutrient solution, which may have affected the K absorption. Blevins et al. (1974) studied the effects of NO_3^- on cation uptake. Fruit K reduction is also known to be related to the harvesting period. The lowest level of K in fruit was measured when harvesting at the end of the harvesting period (Table 6). It is possible that at this end stage plants take up less K from the nutrient solution and it accumulates at a low level in fruit. Na and Ca concentrations of strawberry fruits significantly varied with different treatments (Table 6). Asao et al. (2013) also found that a reduced level of KNO_3 in the nutrient solution increased the Na content in melon fruit. A reduced level of KNO₃ with Ca(NO₃)₂ adjustment increased the Ca contents of mid-and final-stage fruit and the Na content in mid-stage fruit (Table 6). This suggested that the increment of these ions compensated for the reduction in K.

A reduced level of K in nutrient solution was shown to lead to a drastic reduction in K accumulation in strawberry plant parts. When strawberry plants were grown with reduced KNO₃, K concentrations in leaves, crowns, and roots significantly decreased (Figs. 4–6; Table 7). These results indicated that strawberry plants grown with a nutrient solution containing a higher level of KNO₃ had a higher K concentration in plant parts at harvest, as we restricted the KNO₃ supply from formation to the anthesis stage, and therefore reduced the amount of K that translocated from the plant parts to fruit during the maturation stage.

Literature Cited

- Albregts, E. E., G. J. Hochmuth, C. K. Chandler, J. Cornell and J. Harrison. 1996. Potassium fertigation requirements of dripirrigated strawberry. J. Amer. Soc. Hort. Sci. 121: 164–168.
- Asao, T., M. Asaduzzaman, M. F. Mondal, M. Tokura, F. Adachi, M. Ueno, M. Kawaguchi and T. Ban. 2013. Impact of reduced potassium nitrate concentrations in nutrient solution on the growth, yield and fruit quality of melon in hydroponics. Sci. Hortic. 164: 221–231.
- Blevins, D. G., A. J. Hiatt and R. H. Lowe. 1974. Influence of nitrate and chloride uptake on expressed sap pH, organic acid

synthesis, and potassium accumulation in higher plants. Plant Physiol. 54: 82-87.

- Britto, D. T. and H. J. Kronzucker. 2008. Cellular mechanisms of potassium transport in plants. Physiol. Plant. 133: 637–650.
- Cakmak, I. 2005. The role of potassium in alleviating detrimental effects of abiotic stresses in plants. J. Plant Nutr. Soil Sci. 168: 521–530.
- Choi, H. Y. and S. K. Ha. 2013. Potassium balances in maintenance hemodialysis. Electrolyte Blood Press. 11: 9–16.
- Committee for Revision of Diet Therapy Guideline. 2007. Dietary recommendations for chronic kidney disease 2007. Jpn. J. Nephrol. 49: 871–878 (In Japanese).
- Crawford, A. and H. Harris. 2011. Balancing act: Na⁺ sodium K⁺ potassium. Nursing 41: 44–50.
- Ebelhar, S. A. and E. C. Varsa. 2000. Applications in sustainable production: Tillage and potassium placement effects on potassium utilization by corn and soybean. Commun. Soil Sci. Plant Anal. 31: 2367–2377.
- Egilla, J. N., F. T. Davies and M. C. Drew. 2001. Effects of potassium on drought resistance of *Hibiscus rosa-sinensis* cv. Leprecham: Plant growth, leaf macro and micro nutrient content and root longevity. Plant Soil 229: 213–224.
- Hassanloui, M. R. D., M. Taheri and M. J. Malakouti. 2004. The interactive effects of potassium and calcium on the K/Ca and quality of apple fruits (in Naghadeh). J. Agr. Eng. Res. 5: 71–84.
- He, J., K. T. Mills, L. J. Appel, W. Yang, J. Chen, S. E. Rosas, A. Porter, G. Makos, M. R. Weir, L. L. Hmm and J. Kusek. 2014. Urinary sodium and potassium excretion and progression of chronic kidney disease. Circulation 129 (Suppl. 1): 294.
- Hochmuth, G., K. Shuler, E. Hanlon and N. Roe. 1994. Pepper response to fertilization with soluble and controlled release potassium fertilizers. Proc. Fla. State Hort. Soc. 107: 132– 139.
- Holbrook, J. T., K. Y. Patterson, J. E. Bodner, L. W. Douglas, C. Veillon, J. L. Kelsay, W. Mertz and J. C. Smith. 1984. Sodium and potassium intake and balance in adults consuming self selected diets. Am. J. Clin. Nutr. 40: 786–793.
- Hori, H. 1966. Gravel culture of vegetables and ornamentals (In Japanese). Yokendo, Tokyo.
- Imai, E., M. Horio, K. Iseki, K. Yamagata, T. Watanabe, S. Hara, N. Ura, Y. Kiyohara, H. Hirakata, T. Moriyama, Y. Ando, K. Nitta, D. Inaguma, I. Narita, H. Iso, K. Wakai, Y. Yasuda, Y. Tsukamoto, S. Ito, H. Makino, A. Hishida and S. Matsuo. 2007. Prevalence of chronic kidney disease (CKD) in the Japanese general population predicted by the MDRD equation modified by a Japanese coefficient. Clin. Exp. Nephrol. 11: 156–163.
- Jordan-Meille, L. and S. Pellerin. 2004. Leaf area establishment of a maize (*Zea mays L.*) field crop under potassium deficiency. Plant Soil 265: 75–92.
- Kanai, S., K. Ohkura, J. J. Adu-Gyamfi, P. K. Mohapatra, N. T. Nguyen, H. Saneoka and K. Fujita. 2007. Depression of sink activity precedes the inhibition of biomass production in tomato plants subjected to potassium deficiency stress. J. Exp. Bot. 58: 2917–2928.
- Kes, P. 2001. Hyperkalemia: A potentially lethal clinical condition. Acta Clin. Croat. 40: 215–225.
- Koshikawa, K. and M. Yasuda. 2003. Studies on the bench culture with closed hydroponic system in strawberry (Part I). J. Japan. Soc. Hort. Sci. 72 (Suppl. 2): 394 (In Japanese).
- Leigh, R. A. and R. G. Wyn Jones. 1984. A hypothesis relating critical potassium concentrations for growth to the distribution and function of this ion in the plant cell. New Phytol.

97: 1-13.

- Lester, G. E., J. L. Jifon and D. J. Makus. 2006. Supplemental foliar potassium applications with or without a surfactant can enhance netted muskmelon quality. HortScience 41: 741– 744.
- Lester, G. E., J. L. Jifon and G. Rogers. 2005. Supplemental foliar potassium applications during muskmelon fruit development can improve fruit quality, ascorbic acid, and beta-carotene contents. J. Amer. Soc. Hort. Sci. 130: 649–653.
- Locascio, S. J. and G. J. Hochmuth. 2002. Water melon production as influenced by lime, gypsum, and potassium. HortScience 37: 322–324.
- Miller, A. J. and M. D. Cramer. 2004. Root nitrogen acquisition and assimilation. Plant Soil 274: 1–36.
- Mills, K. T., J. Chen, W. Yang, L. Appel, J. Kusek, A. B. Alper, P. Delafontaine, M. G. Keane, E. R. Mohler, A. O. Ojo, M. Rahman, A. C. Ricardo, E. Z. Soliman, S. Steigerwalt, R. R. Townsend and J. He. 2014. Urinary sodium and potassium excretion and cardiovascular diseases in patients with chronic kidney disease: the chronic renal insufficiency cohort study. Circulation 129 (Suppl. 1): 188.
- Mullins, G. L. and C. H. Burmester. 1998. Potassium uptake by crops during the season. p. 123–132. In: D. M. Oosterhuis and G. A. Berkowitz (eds.). Frontiers in Potassium Nutrition: New Perspectives on the Effects of Potassium on Physiology of Plants. Potash and Phosphate Institute, Norcross, GA/Potash and Phosphate Institute of Canada, Saskatoon, Canada.
- Ogawa, A., S. Taguchi and C. Kawashima. 2007. A cultivation method of spinach with a low potassium content for patients on dialysis. Jpn. J. Crop Sci. 76: 232–237 (In Japanese with English abstract).
- Perkins-Veazie, P., W. Roberts and K. Perez. 2003. Water melon fruit potassium and lycopene content in response to increased potassium fertility. HortScience 8: 816–817.
- Pettersson, S. and P. Jensen. 1983. Variation among species and varieties in uptake and utilization of potassium. Plant Soil 72: 231–237.
- Putcha, N. and M. Allon. 2007. Management of hyperkalemia in dialysis patients. Semin. Dial. 20: 431–439.

- Rebolledo-Martinez, A., A. Lid-del-Angel-Perez and J. Rey-Moreno. 2008. Effects of paclobutrazol and KNO₃ over flowering and fruit quality in two cultivars of mango Manila. Interciencia 33: 518–522.
- Rengel, Z. and P. M. Damon. 2008. Crops and genotypes differ in efficiency of potassium uptake and use. Physiol. Plant. 133: 624–636.
- Russell, S. S. 2009. Fluid/electrolyte/acid-base imbalances. p. 116–125. In: H. Craven (ed.). Core Curriculum for Medical-Surgical Nursing. 4th ed. Academy of Medical Surgical Nursing, Pitman, NJ.
- Singh, K. N. and D. P. Sharma. 2001. Response of wheat to nitrogen and potassium in saline soils. Exp. Agric. 37: 417–427.
- Smyth, A., D. Dunkler, P. Gao, K. K. Teo, S. Yusuf, M. J. O'Donnell, J. F. E. Mann and C. M. Clase. 2014. The relationship between estimated sodium and potassium excretion and subsequent renal outcomes. Kidney International 86: 1205–1212.
- Standard Tables of Food Composition in Japan. 2011. Standard Tables of Food Composition in Japan. 5th and enlarged ed. Kagawa Nutrition University Publishing Division, Tokyo.
- Szczerba, M. W., D. T. Britto and H. J. Kronzucker. 2009. K⁺ transport in plants: physiology and molecular biology. J. Plant Physiol. 166: 447–466.
- Umamaheswarappa, P. and K. S. Krishnappa. 2004. Effect of nitrogen, phosphorus and potassium on cucumber cv. Poinsette grown in dry region of southern India. Trop. Sci. 44: 174– 176.
- US Renal Data System. 2002. USRDS 2002 Annual Data Report. The National Institutes of Health, National Institutes of Diabetes and Digestive and Kidney Diseases, Bethesda, MD.
- Voogt, W. and C. Sonneveld. 1997. Nutrient management in closed growing systems for greenhouse production. p. 83– 102. In: E. Goto (ed.). Plant Production in Closed Ecosystem. Academic Publishers, Dordrecht.
- Woodend, J. J. and A. D. M. Glass. 1993. Genotype environment interaction and correlation between vegetative and grain production measures of potassium use efficiency in wheat (*T. aestivum* L.) grown under potassium stress. Plant Soil 5: 39–44.